

LSBio™ Mouse F10 / Factor X ELISA Kit

Catalog No. LS-F10442

User Manual

**Please Read the Manual Carefully
Before Starting your Experiment**



For research use only. Not approved for use in humans or for clinical diagnosis.

INTENDED USE

This mouse Factor X total assay is for the quantitative determination of total Factor X and Xa in biological fluids.

For research use only.

BACKGROUND

Factor X is a disulfide linked two-chain glycoprotein zymogen and is the precursor of the coagulation enzyme Factor Xa [1]. Factor X serves as the intersection of the intrinsic and extrinsic coagulation cascades and can be activated by either the extrinsic Factor VIIa / Tissue Factor complex or the intrinsic Factor IXa / Factor VIIIa complex. Factor Xa converts prothrombin to thrombin and is quickly inhibited by Antithrombin III in the presence of heparin.

ASSAY PRINCIPLE

Mouse Factor X will bind to the affinity purified capture antibody coated on the microtiter plate. Factor X, Xa, and Xa in complex with inhibitors will react with the antibody on the plate. After appropriate washing steps, biotin labeled polyclonal anti-mouse Factor X primary antibody binds to the Factor X. Excess antibody is washed away and bound polyclonal antibody is then reacted with Streptavidin conjugated to horseradish peroxidase. Following an additional washing step, TMB substrate is used for color development at 450nm. The amount of color development is directly proportional to the concentration of total Factor X in the sample.

REAGENTS PROVIDED

- **96-well antibody coated microtiter strip plate** (removable wells 8x12) containing affinity purified sheep anti-mouse Factor X antibody, blocked and dried.
- **10X Wash Buffer:** 1 bottle of 50ml
- **Mouse Factor X standard:** 1 vial lyophilized standard
- **Anti-Factor X primary antibody:** 1 vial lyophilized biotin labeled polyclonal antibody
- **Horseradish peroxidase-conjugated Streptavidin:** 1 vial concentrated HRP labeled streptavidin
- **TMB substrate solution:** 1 bottle of 10ml solution

STORAGE AND STABILITY

Store all kit components at 4°C upon arrival. Return any unused microplate strips to the plate pouch with desiccant. Reconstituted standards and primary may be stored at -80°C for later use. Do not freeze-thaw the standard and primary antibody more than once. Store all other unused kit components at 4°C. This kit should not be used beyond the expiration date.

OTHER REAGENTS AND SUPPLIES REQUIRED

- Microtiter plate shaker capable of 300 rpm uniform horizontally circular movement
- Manifold dispenser/aspirator or automated microplate washer
- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and Pipette tips
- Deionized or distilled water
- Polypropylene tubes for dilution of standard
- Paper towels or laboratory wipes
- 1N H₂SO₄ or 1N HCl
- Bovine Serum Albumin Fraction V (BSA)
- Tris(hydroxymethyl)aminomethane (Tris)
- Sodium Chloride (NaCl)

PRECAUTIONS

- **FOR LABORATORY RESEARCH USE ONLY. NOT FOR DIAGNOSTIC USE.**
- Do not mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
- Always pour peroxidase substrate out of the bottle into a clean test tube. Do not pipette out of the bottle as contamination could result.
- Keep plate covered except when adding reagents, washing, or reading.
- DO NOT pipette reagents by mouth and avoid contact of reagents and specimens with skin.
- DO NOT smoke, drink, or eat in areas where specimens or reagents are being handled.

PREPARATION OF REAGENTS

- TBS buffer:** 0.1M Tris, 0.15M NaCl, pH 7.4
- Blocking buffer (BB):** 3% BSA (w/v) in TBS
- 1X Wash buffer:** Dilute 50ml of 10X wash buffer concentrate with 450ml of deionized water.

SAMPLE COLLECTION

Collect plasma using citrate as an anticoagulant. Heparin or EDTA is not recommended. Heparin binds Factor X thus interfering with the assay. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. Assay immediately or aliquot and store at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

ASSAY PROCEDURE

Perform assay at room temperature. Vigorously shake plate (300rpm) at each step of the assay.

Preparation of Standard

Reconstitute standard by adding 1ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. This will result in a 1,000ng/mL standard solution.

Dilution table for preparation of mouse Factor X standard:

Factor X concentration ng/ml	Dilutions
500	500 μl (BB) + 500 μl (1000ng/ml)
200	600 μl (BB) + 400 μl (500ng/ml)
100	500 μl (BB) + 500 μl (200ng/ml)
50	500 μl (BB) + 500 μl (100ng/ml)
25	500 μl (BB) + 500 μl (50ng/ml)
10	600 μl (BB) + 400 μl (20ng/ml)
5	500 μl (BB) + 500 μl (10ng/ml)
2.5	500 μl (BB) + 500 μl (5ng/ml)
0	500 μl (BB) Zero point to determine background

NOTE: DILUTIONS FOR THE STANDARD CURVE AND ZERO STANDARD MUST BE MADE AND APPLIED TO THE PLATE IMMEDIATELY.

Standard and Unknown Addition

Remove microtiter plate from bag and add 100 μl Factor X standards (in duplicate) and unknowns to wells. Carefully record position of standards and unknowns. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300 μl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

NOTE: The assay measures Factor X antigen in the 2.5-500 ng/ml range. If the unknown is thought to have high Factor X levels, dilutions may be made in blocking buffer. A 1:1,000-1:2,000 dilution for normal mouse plasma or serum is suggested for best results.

Primary Antibody Addition

Reconstitute primary antibody by adding 10ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. Add 100 μl to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300 μl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Streptavidin-HRP Addition

Dilute 2.5 μl of HRP conjugated streptavidin into 2.5ml diluent to generate a 1:1,000 dilution. Add 0.2ml of 1:1,000 dilution to 9.8ml of diluent to generate a 1:50,000 dilution. Add 100 μl of the 1:50,000 dilution to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300 μl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Substrate Incubation

Add 100 μl TMB substrate to all wells and shake plate for 2-7 minutes. Substrate will change from colorless to different strengths of blue. Quench reaction by adding 50 μl of 1N H_2SO_4 or HCl stop solution to all wells when samples are visually in the same range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

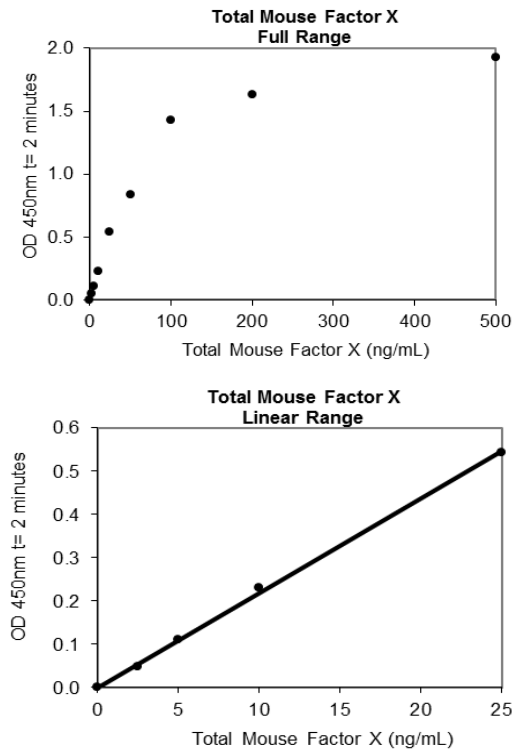
Measurement

Set the absorbance at 450nm in a microtiter plate spectrophotometer. Measure the absorbance in all wells at 450nm. Subtract zero point from all standards and unknowns to determine corrected absorbance (A_{450}).

Calculation of Results

Plot A_{450} against the amount of Factor X in the standards. Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve. Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit. The amount of Factor X in the unknowns can be determined from this curve. If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

A typical standard curve (EXAMPLE ONLY):



EXPECTED VALUES

The concentration of Factor X in normal human plasma was found to be 10 µg/ml [2]. Normal values of Factor X

in mouse plasma have not been conclusively determined but are believed to be similar to human plasma. Oral anticoagulants such as warfarin reduce functional Vitamin K and disrupt the post-translational addition of gamma-carboxyglutamic acid (gla) residues, decreasing the thrombotic activity of Factor Xa but not the concentration of Factor X antigen (3).

PERFORMANCE CHARACTERISTICS

Sensitivity: The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates (range OD₄₅₀: 0.069-0.075) and calculating the corresponding concentration. The MDD was 0.146 ng/ml.

Intra-assay Precision: These studies are currently in progress. Please contact us for more information.

Inter-assay Precision: These studies are currently in progress. Please contact us for more information.

Recovery: These studies are currently in progress. Please contact us for more information.

Linearity: These studies are currently in progress. Please contact us for more information.

Specificity: These studies are currently in progress. Please contact us for more information.

Sample Values: Samples were evaluated for the presence of the antigen at varying dilutions.

Sample Type	Dilution	Mean (µg/ml)
Citrate Plasma	1:1,000	22.7
	1:2,000	23.1

DISCLAIMER

This information is believed to be correct but does not claim to be all-inclusive and shall be used only as a guide. The supplier of this kit shall not be held liable for any damage resulting from handling of or contact with the above product.

REFERENCES

1. Di Scipio RG, *et al.*: Biochemistry. 1977, 16(4): 698-706.
2. Berthier AM, *et al.*: Haemostasis. 1982. 142. 3.
- Kumar S, *et al.*: Br J Haematol. 1990. 74(1): 82-5.

Example of ELISA Plate Layout

96 Well Plate: 18 Standard wells, 78 Sample wells

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	2.5 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	200 ng/ml	500 ng/ml			
B	0	2.5 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	200 ng/ml	500 ng/ml			
C												
D												
E F												
G												
H												

Important Note: During shipment, small volumes of product will occasionally become entrapped in the seal of the product vial. We recommend briefly centrifuging the vial to dislodge any liquid in the container's cap prior to opening.

Warning: This reagent may contain sodium azide and sulfuric acid. The chemical, physical, and toxicological properties of these materials have not been thoroughly investigated. Standard Laboratory Practices should be followed. Avoid skin and eye contact, inhalation, and ingestion. Sodium azide forms hydrazoic acid under acidic conditions and may react with lead or copper plumbing to form highly explosive metal azides. On disposal, flush with large volumes of water to prevent accumulation.

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